



Seasonal variation of aflatoxin M₁ contamination in industrial and traditional Iranian dairy products

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ABSTRACT

This study aimed to determine the occurrence of aflatoxin M₁ (AFM₁) contamination in 682 dairy product samples consisting of raw milk of cow, goat and sheep; Lighvan cheese; and industrial and traditional yoghurt, Kashk and Doogh samples collected from popular markets and dairy ranches in four large Iranian cities. Thin layer chromatography (TLC) technique was used for analysis of the samples. Results showed that the incidence and levels of AFM₁ contamination in raw cow milk and industrial products (manufactured from cow milk) were higher than raw goat or sheep milk, and traditional products (made from goat and sheep milk), respectively. Moreover, seasonal variations influenced the concentration of AFM₁ in most of the analyzed dairy products. Owing to the abundance and popularity of the industrial products, contamination of these products in such a level could be a potential hazard for public health.

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1. Introduction

Aflatoxins are a group of extremely toxic metabolites of fungi produced by toxicogenic strains of *Aspergillus flavus*, *Aspergillus parasiticus* and *Aspergillus nomius* in a wide variety of agricultural commodities (Mortazavi & Tabatabai, 1998). They are highly toxic, carcinogenic, mutagenic and teratogenic compounds that have been considered as causative agent in human hepatic and extra-hepatic carcinogenesis. Four common types of aflatoxins are B₁, B₂, G₁ and G₂. Among them, aflatoxin B₁ (AFB₁) is notoriously the most frequent produced mycotoxin; and has been reported to be the most powerful natural carcinogen in mammals (Creppy, 2002; Hussain, Anwar, Munawar, & Asi, 2008; Tokar & Vengust, 2008). The International Agency for Research on Cancer (IARC, 1993) of WHO included AFB₁ as Group 1 human carcinogen.

Aflatoxin M₁ (AFM₁) is the monohydroxylated metabolite of AFB₁, metabolized by cytochrome P450 enzyme system in liver and

excreted into the milk of lactating livestock which consumed AFB₁ contaminated diet (Gürbay, Aydın, Girgin, Engin, & Şahin, 2006; Keskin, Başkaya, Karsli, Yurdun, & Özyaral, 2009). There is a direct relationship between the AFM₁ content in milk and AFB₁ consumption via foodstuffs. It has been estimated that about 0.5–6% of AFB₁ present in animal feed pass as AFM₁ in milk. This variability is due to different factors such as stage and order of lactation, intake level of AFB₁ and individual response (Galvano, Galofaro, & Galvano, 1996). The toxicity of AFM₁ is less than its parent compound, AFB₁; but the cytotoxic, genotoxic, and carcinogenic effects of it is well demonstrated. Hence, the IARC of WHO re-considered its carcinogenicity classification and changed it from Group 2 to Group 1 according to more recent investigations (IARC, 2002).

When dairy products are manufactured from AFM₁ contaminated milk, the toxin could be detected in them (Bakirci, 2001). Unfortunately, the content of AFM₁ remains relatively stable during processing and storage of various dairy products. This toxin cannot be inactivated by thermal processing used in dairy industry i.e. pasteurization and ultra-high-temperature (UHT) treatment (Fallah, 2010a,b).

Several countries have legislated permissible levels of AFM₁ in milk and dairy products to protect consumers specially children. These regulations are influenced by economic considerations and vary from one country to another (Fallah, 2010b). The US Food and

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Drug Administration (US FDA, 1996, p. 219) has set a limit of 0.5 µg/l for AFM₁ in milk. However, the Institute of Standards and Industrial Research of Iran (ISIRI, 2002) has accepted 0.050 µg/l as the action level for AFM₁ which is the same as European Commission permitted level (European Commission, 2001). Additionally, according to ISIRI (2002), the maximum admissible level of AFM₁ in other dairy products is 0.5 µg/kg in dried milk, 0.020 µg/kg in butter and buttermilk, 0.250 µg/kg in cheese and Kashk; and 0.050 µg/l in yoghurt and Doogh.

Several studies in the world have been undertaken to determine the occurrence of AFM₁ in milk and dairy products (Carvajal, Bolaños, Rojo, & Méndez, 2003; Elzupir & Elhussein, 2010; Ghanem & Orfi, 2009; Hampikyan, Bingol, Cetin, & Colak, 2010; Kim, Lee, Kwak, Ahn, & Jeong, 2010; Nuryono et al., 2009). However, referring to existing scientific literature, a limited number of surveys have been conducted in this field in Iran. Furthermore, no seasonal study on the levels of AFM₁ in goat milk, sheep milk and traditional dairy products has been performed in Iran. Therefore, this study aimed to determine the occurrence of AFM₁ in industrial and traditional dairy products widely marketed all over the country as well as raw milk during different seasons.

2. Materials and methods

2.1. Sample collection

A total of 682 dairy product samples composed of raw cow milk ($n = 88$), raw goat milk ($n = 65$), raw sheep milk ($n = 72$), Lighvan cheese ($n = 75$), industrial yoghurt ($n = 61$), traditional yoghurt ($n = 60$), industrial Kashk ($n = 64$), traditional Kashk ($n = 61$), industrial Doogh ($n = 71$) and traditional Doogh ($n = 65$) were obtained from dairy ranches, supermarkets and retail outlets in four large Iranian cities (Tehran, Esfahan, Tabriz and Shiraz) during year 2008. The samples were transported to the laboratory inside a digital portable refrigerator at 3 °C and stored at –20 °C until analysis for AFM₁.

2.2. Reagents

All reagents were analytically pure and purchased from Merck KgaA (Darmstadt, Germany). AFM₁ standard was obtained from Sigma Chemical Company, USA. Stock solution of AFM₁ (50 mg/ml) was prepared in a methanol/chloroform mixture (81:19, v/v) and stored at –20 °C. Before use, the stock solution was diluted with methanol/chloroform (1/1, v/v) at appropriate concentrations (Fallah, 2010a,b).

2.3. Apparatus

2.3.1. Thin layer plates

Silica gel 60 (Merck, dimensions 20 × 20 cm², thickness 0.2 mm). These plates were cut in 10 × 10 cm² dimensions and used for the experiment.

2.3.2. Ultraviolet lamp

A Fisher ultraviolet (UV) lamp with 245 and 364 nm wavelengths was obtained from Philips.

2.4. Methods

Samples of milk and dairy products were analyzed by two-dimensional thin layer chromatography (TLC) technique for presence of AFM₁ according to the method of Stubblefield (1979) and Karim (1998) as described in the previous paper (Fallah, 2010a).

A homogenized sample (50 ml or 50 g) was mixed with 10 ml of saturated sodium chloride (cooled at 4 °C) and 125 ml of chloroform (cooled at 4 °C) for 1.5 min. The mixture was transferred into a 250-ml centrifuge bottle and centrifuged at 3700 rpm, in order to separate phases. The lower phase (chloroform layer) was drained into 300-ml Erlenmeyer flask containing 5 g of anhydrous sodium sulphate and swirled intermittently for 15 min. Then the mixture was filtered through a Whatman No. 4 filter paper into a 250-ml graduated cylinder to obtain sample filtrate and the volume was recorded. The filtrate was purified by silica gel column chromatography and then submitted to solvent evaporation with a rotary vacuum evaporator until nearly dry. Subsequently, the residue was dissolved in ~4 ml of chloroform and dried by a weak stream of nitrogen at 50 °C. For AFM₁ assay, the dried extract was resuspended in 100 µl of chloroform. Thereafter, 25 µl of the sample extract and a series of 3, 6, 12 and 24 µl of standard AFM₁ working solution (0.05 µg/ml) were spotted on the thin layer plate of silica gel. The plate was developed in the first and second directions with diethyl ether:methanol:water (94:4.5:1.5, v/v/v) and chloroform:acetone:methanol (87:10:3, v/v/v), respectively. The concentration of AFM₁ in the sample was estimated by visual comparison of sample extract fluorescent spot with that of standard AFM₁ spots under long wave UV light (364 nm); using the formula described in the previous paper (Fallah, 2010a). The confirmatory test for AFM₁ was carried out by adding trifluoroacetic acid to the fluorescent spots and comparing the thin layer chromatographic properties of the derivatives of the sample spot and the standard. The lowest detection limit of the method is 0.0125 ppb.

2.5. Method evaluation

In order to validate our method, homogenized samples of raw milk, Lighvan cheese, yoghurt, Kashk and Doogh were spiked with AFM₁ at levels of 0.05, 0.1 and 0.2 ppb just before the test. The AFM₁ concentration was determined using the previously described procedure. The results indicated that the method was reliable for determination of AFM₁ in the mentioned dairy products (Table 1).

2.6. Statistical analyses

To evaluate seasonal effects, data obtained from each dairy product sample in different seasons were compared. Statistical analyses were carried out using one-way ANOVA of the GraphPad Prism software, version 3. The differences among means at $P < 0.05$ were compared by Tukey's Multiple Comparison Test.

Table 1

Validation of the TLC method for determination of AFM₁ in dairy products ($n = 4$).

Dairy product	Spiked level (ppb)	AFM ₁ determined (ppb)	Recovery ^a (%)	Coefficient of variation (%)
Raw milk	0.05	0.045	90.5 ± 4.9	5.4
	0.10	0.088	87.6 ± 5.8	6.6
	0.20	0.175	87.5 ± 8.3	9.5
Lighvan cheese	0.05	0.040	79.8 ± 9.1	11.3
	0.10	0.088	88.5 ± 7.9	9.0
	0.20	0.175	87.3 ± 9.1	10.4
Yoghurt	0.05	0.044	87.6 ± 6.8	7.8
	0.10	0.084	83.5 ± 8.5	10.1
	0.20	0.159	79.7 ± 4.8	6.1
Kashk	0.05	0.038	75.5 ± 7.0	9.3
	0.10	0.083	82.5 ± 9.4	11.4
	0.20	0.166	83.0 ± 8.2	9.8
Doogh	0.05	0.046	92.5 ± 4.5	4.8
	0.10	0.086	85.7 ± 8.1	9.5
	0.20	0.179	89.4 ± 7.0	7.8

^a Mean ± SD.

Table 2
Occurrence of aflatoxin M₁ in dairy products from Iran.

Dairy product	Samples analyzed, <i>n</i>	Positive samples, <i>n</i> (%)	Mean ± SEM (μg/l or μg/kg) ^a	Min–max (μg/l or μg/kg) ^a	Exceed regulation, <i>n</i> (%) ^b
Raw cow milk	88	74 (84.1)	0.052 ± 0.006	0.013–0.394	31 (35.2)
Raw goat milk	65	28 (43.1)	0.018 ± 0.003	0.013–0.055	7 (10.8)
Raw sheep milk	72	43 (59.7)	0.027 ± 0.004	0.015–0.102	18 (25.0)
Lighvan cheese	75	49 (65.3)	0.085 ± 0.010	0.030–0.313	7 (9.3)
Industrial yoghurt	61	30 (49.2)	0.026 ± 0.004	0.015–0.102	10 (16.4)
Traditional yoghurt	60	14 (23.3)	0.007 ± 0.002	0.015–0.036	0 (0.0)
Industrial Kashk	64	34 (53.1)	0.080 ± 0.013	0.028–0.285	11 (17.2)
Traditional Kashk	61	19 (31.2)	0.053 ± 0.011	0.046–0.291	1 (1.6)
Industrial Doogh	71	16 (22.5)	0.007 ± 0.002	0.013–0.053	3 (4.2)
Traditional Doogh	65	9 (13.8)	0.003 ± 0.001	0.013–0.029	0 (0.0)

^a Results are expressed in μg/l for raw milk and Doogh; and μg/kg for cheese, yoghurt and Kashk.

^b The ISIRI limit for aflatoxin M₁ is 0.050 ppb for milk, yoghurt and Doogh; and 0.250 ppb for cheese and Kashk.

3. Results and discussion

3.1. Raw milk

In the present study, the incidence of AFM₁ contamination in raw cow, goat and sheep milk was 84.1%, 43.1% and 59.7%; with the average values of 0.052, 0.018 and 0.027 μg/l, respectively. However, the levels of the toxin in 35.2% of cow milk, 10.8% of goat milk and 25% of sheep milk samples exceeded the ISIRI and EC limit for AFM₁ in liquid milk i.e. 0.050 μg/l (Table 2). The lower concentrations of AFM₁ in goat and sheep milk than cow milk could be due to the pasture grazing of goat and sheep in Iran. It has been demonstrated that out-pasturing is an effective factor to decrease the level of AFM₁ contamination in milk of dairy species (Hussain, Anwar, Asi, Munawar, & Kashif, 2010; Kamkar, 2005). Only for 2–3 months in winter, Iranian goat and sheep are fed by stored grains and compound feed. The obtained results are comparable to the results of previous studies performed in Iran (Ghiasian, Maghsood, Neyestani, & Mirhendi, 2007; Kamkar, 2005; Rahimi, Bonyadian, Rafei, & Kazemeini, 2010; Tajkarimi et al., 2008); but are higher than those reported in European countries (Bilandžić, Varenina, & Solomun, 2010; Martins & Martins, 2000; Saccà et al., 2009; Velasco, Delso, & Escudero, 2003; Virdis, Corgiolu, Scarano, Pilo, & De Santis, 2008). This is the consequence of comprehensive supervision and Hazard Analysis and Critical Control Point (HACCP) programmes developed in European countries to manage the risks associated with toxicogenic fungi contamination all along the feed supply chain; and also legislating strict regulatory limits for aflatoxins in feed and milk.

Considering seasonal effect influences, the levels of AFM₁ in cow, goat and sheep milk samples collected in winter were significantly higher ($P < 0.05$) than those collected in the other seasons (Table 3).

Our findings are in agreement with previous studies that reported higher levels of AFM₁ contamination in cold seasons than hot ones (Hussain & Anwar, 2008; Nemati, Mehran, Hamed, & Masoud, 2010; Ruangwises & Ruangwises, 2010). The reason is that in cold seasons lactating animals are fed with greater amounts of compound feed which may be contaminated with higher levels of AFB₁.

3.2. Lighvan cheese

Lighvan cheese is a semi-hard, starter-free, traditional Iranian cheese produced from a mixture of raw goat and sheep milk in small dairies located in Lighvan village in East Azarbaijan province of Iran and widely consumed all over the country. The frequency of AFM₁ contamination in Lighvan cheese was 65.3%, at the mean value of 0.085 μg/kg; and 9.3% had levels of the toxin above the legal limit accepted by ISIRI i.e. 0.250 μg/kg (Table 2). Due to the preferential affinity of AFM₁ for casein fraction of milk, concentration of the toxin is higher in cheese than in milk from which the cheese is made (Ardic, Karakaya, Atasever, & Adiguzel, 2009). Referring to the existing scientific literature, no data have been published on the occurrence of AFM₁ in cheese made from goat milk, sheep milk or mixture of them in Iran. Comparing our findings with the previous studies conducted in Iran (Fallah, 2010a; Fallah, Jafari, Fallah, & Rahnama, 2009; Kamkar, 2006), this survey revealed lower AFM₁ contamination in Lighvan cheese than white cheese prepared from cow milk.

Considering seasonal variability, the mean concentration of AFM₁ in Lighvan cheese samples collected in spring was significantly higher ($P < 0.05$) than those collected in the other seasons (Table 3). Due to the duration of ripening of Lighvan cheese (3 months), most of the cheeses offered for sale in spring are manufactured from winter milk that could contain higher levels of AFM₁.

Table 3
Levels of aflatoxin M₁ in dairy products from Iran: Comparison between samples obtained in different seasons.

Dairy product	Winter		Spring		Summer		Autumn	
	Samples analyzed, <i>n</i>	Mean ± SEM (μg/l or μg/kg) ^d	Samples analyzed, <i>n</i>	Mean ± SEM (μg/l or μg/kg) ^d	Samples analyzed, <i>n</i>	Mean ± SEM (μg/l or μg/kg) ^d	Samples analyzed, <i>n</i>	Mean ± SEM (μg/l or μg/kg) ^d
Raw cow milk	24	0.093 ± 0.019 ^a	21	0.031 ± 0.005 ^c	21	0.028 ± 0.005 ^c	22	0.051 ± 0.006 ^b
Raw goat milk	18	0.035 ± 0.005 ^a	17	0.013 ± 0.004 ^b	16	0.011 ± 0.004 ^b	14	0.012 ± 0.005 ^b
Raw sheep milk	22	0.046 ± 0.008 ^a	19	0.021 ± 0.007 ^b	15	0.019 ± 0.005 ^b	16	0.019 ± 0.006 ^b
Lighvan cheese	20	0.083 ± 0.020 ^b	17	0.162 ± 0.024 ^a	20	0.051 ± 0.015 ^b	18	0.053 ± 0.018 ^b
Industrial yoghurt	16	0.038 ± 0.010 ^a	15	0.013 ± 0.006 ^b	16	0.012 ± 0.005 ^b	14	0.041 ± 0.009 ^a
Traditional yoghurt	15	0.018 ± 0.004 ^a	15	0.002 ± 0.001 ^b	15	0.002 ± 0.002 ^b	15	0.005 ± 0.003 ^b
Industrial Kashk	15	0.115 ± 0.025 ^a	18	0.130 ± 0.026 ^a	17	0.039 ± 0.017 ^b	14	0.030 ± 0.018 ^b
Traditional Kashk	15	0.047 ± 0.024	16	0.051 ± 0.022	15	0.066 ± 0.026	15	0.053 ± 0.011
Industrial Doogh	18	0.007 ± 0.004	16	0.008 ± 0.005	17	0.006 ± 0.003	20	0.009 ± 0.004
Traditional Doogh	16	0.006 ± 0.003	16	0.002 ± 0.002	17	0.000 ± 0.000	16	0.004 ± 0.002

^{a,b,c} Mean ± SEM in the same row with different letters are significantly different ($P < 0.05$).

^d Results are expressed in μg/l for raw milk and Doogh; and μg/kg for cheese, yoghurt and Kashk.

3.3. Yoghurt and yoghurt derived products

Industrial yoghurt is manufactured from cow milk in dairy industries, while traditional yoghurt is made from goat milk, sheep milk or mixture of them in ranches or small dairy shops. Our study showed that the incidence and level of AFM₁ in industrial yoghurt were higher than traditional yoghurt due to the higher occurrence of the toxin in cow milk than goat and sheep milk. The level of AFM₁ in 16.4% of the industrial samples was above the legal limit accepted by ISIRI i.e. 0.050 µg/kg; while none of the samples of traditional yoghurt exceeded the limit (Table 2). Regarding seasonal variability, the levels of the toxin in industrial yoghurt samples obtained in autumn and winter were significantly higher ($P < 0.05$) than those obtained in spring and summer. In the case of traditional yoghurt, the level of AFM₁ was significantly higher ($P < 0.05$) in winter than other seasons (Table 3).

Kashk is a special Iranian dairy product prepared by prolonged boiling of yoghurt. Traditional Kashk is a dried product with the shape of conic or cubic balls originated from goat and sheep milk. But, the industrial Kashk is a thick whitish liquid prepared from cow milk. The frequency of AFM₁ contamination in industrial and traditional Kashk was 53.1% and 31.2%, with the average values of 0.080 and 0.053 µg/kg, respectively. It was found that 11 samples (17.2%) of industrial and one sample (1.6%) of traditional Kashk had levels of the toxin in excess of the ISIRI limit i.e. 0.250 µg/kg (Table 2). In the case of industrial Kashk, winter and spring levels of AFM₁ contamination were significantly higher ($P < 0.05$) than summer and autumn levels of the toxin. In contrast, no statistically significant seasonal effect ($P > 0.05$) was observed for traditional Kashk samples (Table 3).

Doogh is a traditional Iranian fermented product prepared from yoghurt by adding potable water, starter culture and sodium chloride. Iranian Doogh is exported and consumed in Afghanistan, Azerbaijan, Armenia, Turkey, Balkans and to a lesser extent in other parts of Middle East and central Asia (Codex Alimentarius Commission, 2009). In the present study the incidence of AFM₁ contamination in Doogh samples was approximately low, since the toxin was detected in 22.5% of industrial Doogh (manufactured from cow milk) and 13.8% of traditional Doogh samples (prepared from goat and sheep milk). The average values of AFM₁ contamination were 0.007 and 0.003 µg/l in industrial and traditional Doogh, respectively; and only 4.2% of the industrial Doogh samples exceeded the ISIRI limit i.e. 0.050 µg/l (Table 2). No statistically significant seasonal effect ($P > 0.05$) was found for Doogh samples obtained in different seasons (Table 3).

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